

(19)



Europäisches Patentamt

European Patent Office

Office européen des brevets



(11)

EP 0 440 619 B1

(12)

EUROPEAN PATENT SPECIFICATION

(45) Date of publication and mention
of the grant of the patent:
24.01.1996 Bulletin 1996/04

(51) Int. Cl.⁵: A61K 39/395, C12N 5/00,
A61K 51/00, A61K 49/00

(21) Application number: 89903537.2

(86) International application number: PCT/US88/03590

(22) Date of filing: 11.10.1988

(87) International publication number: WO 89/06242
(13.07.1989 Gazette 1989/15)

(54) ANTIBODY TO A4 AMYLOID PEPTIDE

ANTIKÖRPER GEGEN A4 AMYLOIDPEPTID

ANTICORPS AGISSANT SUR UN PEPTIDE DE L'AMYLOIDE DE SEQUENCE A4

(84) Designated Contracting States:
AT BE CH DE FR GB IT LI LU NL SE

(74) Representative: Sheard, Andrew Gregory et al
London WC1N 2DD (GB)

(30) Priority: 08.10.1987 US 105751

(56) References cited:
US-A- 4 666 829

(43) Date of publication of application:
14.08.1991 Bulletin 1991/33

(73) Proprietors:
• THE McLEAN HOSPITAL CORPORATION
Belmont, MA 02178 (US)
• THE UNIVERSITY OF ROCHESTER
Rochester, New York 14627 (US)

- BIOCHEM. BIOPHYS. RES. COMM., vol 145, pp. 241-248, W.M. PARDRIDGE et al., 29 May 1987
- PROC. NAT. ACAD. SCI. USA, vol. 82, 8729-8732, C.W. WONG et al., 1985
- NATURE, vol. 325, 733-736, J. KANG et al., 19 Feb 1987
- BIOCHEM. BIOPHYS. RES. COMM., vol. 120, 885-890, G. GLENNER et al., 16 May 1984
- PROC. NAT. ACAD. SCI, USA, vol. 84, 4190-4194, ROBAKIS, 1987

(72) Inventors:
• MAJOCHA, Ron
Wayland, MA 01778 (US)
• MAROTTA, Charles, A.
Cambridge, MA 02140 (US)
• ZAIN, Sayeeda

Remarks:

The file contains technical information submitted
after the application was filed and not included in this

Description

BACKGROUND OF THE INVENTION

The present invention was made utilizing funds of the United States Government. The United States Government is hereby granted a worldwide royalty fee, paid up, non-exclusive license to the invention.

FIELD OF THE INVENTION

The present invention relates to an antibody with specificity to A4 amyloid polypeptide found in the brain of Alzheimer's Disease (AD) patients, and to uses thereof, especially for the neuropathological definition of AD senile plaque subtypes, and for the fine, detailed imaging of AD brain tissue.

BRIEF DESCRIPTION OF THE BACKGROUND ART

Although the presence of amyloid in the plaques of Alzheimer's Disease patients was noted over 60 years ago (Divry, 1927), molecular mechanisms that produce amyloid in the aged human brain and for the increased deposition of this fibrous material in Alzheimer's Disease remain unknown. Also undescribed is the contribution, if any, of amyloid to the active process of plaque formation vis-a-vis a secondary and more passive role that indicates only the terminal stages of parenchymal deterioration.

Recently, some progress has been made in defining the partial structure of amyloid fibrillary protein. Glenner *et al.* (1984a) purified amyloid from meningeal vessels of an AD brain; a 4.2 kD polypeptide was isolated and shown to have a unique 24 amino acid sequence (β -polypeptide, Fig. 1). A polypeptide of similar sequence was subsequently isolated from the cerebrovascular amyloid of a Down's syndrome brain (Glenner *et al.*, 1984b); a single amino acid substitution, of glutamic acid for glutamine at position 11, distinguished the two polypeptides. Similar results were independently obtained by Masters *et al.* (1985a) who partly purified and analyzed amyloid plaque cores from the AD cerebral cortex; the 28 amino acid sequence of the Glu variant was obtained (A4 sequence, Fig. 1). The A4 sequence differs from the β -polypeptide disclosed by Glenner *et al.* (USP 4,666,829) by the three amino acids at positions 11, 27 and 28. The significance of this report is that the A4 was derived from amyloid plaque cores, a hallmark feature of AD. The amino acid sequence of A4 varies from that of the β -polypeptide derived from vascular amyloid (see Fig. 1).

Using polyclonal antisera to a synthetic β -amyloid polypeptide containing residues 1 through 10 (Fig. 1), it was shown that neuritic plaque amyloid shares antigenic determinants with the similar fibrillary lesion of cerebral vessels (Wong *et al.*, 1985). The same antisera failed to detect neurofibrillary tangles (NFTs). By contrast, antiserum raised against residues 1 to 11 of the A4 polypeptide

failed to detect vascular amyloid or neuritic plaques but, rather, exhibited exclusive specificity for the NFT; and antisera to the A4 peptide extending from residues 11 to 23 stained both plaques and vessels (Masters *et al.*, 1985b). Thus, there is precedent to believe that antibodies to the β -peptide and the A4 peptide are not identical with regard to their specificities.

Glenner *et al.*, U.S.P. 4,666, 829 disclose the preparation of antibodies using the first 10 amino acids of the β -amyloid polypeptide (Fig. 1).

SUMMARY OF THE INVENTION

In order to facilitate studies on the molecular mechanisms involved in fibrous protein accumulation in the aged AD demented brain and to provide improved neuropathological aids for the diagnosis of AD subtypes, we have prepared a monoclonal antibody (Mab), to a synthetic amyloid polypeptide with the known 28 amino acid A4 sequence (Masters *et al.*, 1985a, Fig. 1). The Mab was routinely characterized on AD cortical and hippocampal sections, and shown to be useful to carry out an analysis of individual epitopic sites of Alzheimer-type amyloid.

The Mab was specifically and extensively characterized and was used to obtain information on the following: (a) detailed morphological features of plaque amyloid revealed by individual target epitopes; (b) the identification of new subtypes of amyloid deposits in the AD brain; (c) the relationship of plaque maturation to the deposition of epitopes. The latter study was made possible through computer-assisted imaging and micro-densitometry.

The antibody can be used in *in vitro* immunoassay procedures for AD-amyloid. It can also be used in imaging (e.g. cytochemical or *in vivo*) neurons for evidence of AD-amyloid. For immunoassays and/or imaging, the antibody can be detectably labelled with, e.g., radio, enzyme or fluorescent labels. It can also be immobilized on insoluble carriers.

The striking aspect of the Mab prepared to the A4 28-mer peptide is that it defines previously undescribed amyloid formations in the AD brain. This Mab represents, therefore, a unique class of Mabs.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 shows the amino acid sequence of the β -polypeptide (Glenner *et al.*, 1984 a,b), the synthetic amyloid A4 peptide (Masters *et al.*, 1985a) used as the antigen in the present invention, and the polypeptide disclosed by Glenner *et al.* (U.S. Patent 4,666,829 (1987)). The difference between the A4 polypeptide and the other two polypeptides are depicted by underlining (amino acids 11, 27 and 28).

DESCRIPTION OF THE PREFERRED EMBODIMENTS

The antibody of the present invention has specificity to one or more epitopes present on the A4 28-mer peptide shown in Figure 1. The antibody of the invention is monoclonal, is made with the 28-mer polypeptide as an immunogen, and can be utilized in the multiple applications described herein below.

The term "epitope" as used in this invention is meant to include any determinant responsible for specific interaction with an antibody molecule. Epitopic determinants usually consist of chemically active surface groupings of molecules such as amino acids or sugar side chains and have specific three dimensional structural characteristics as well as specific charge characteristics.

Polyclonal antibodies can be generated in any suitable animal such as, for example, mice, rabbits or goats. The A4-amyloid 28-peptide can be injected by itself or linked to appropriate immunoactivating carriers, such as KLH. Further detailed descriptions of immunization protocols can be found in the Examples.

Monoclonal antibodies can be produced in various ways using techniques well understood by those having ordinary skill in the art and will not be repeated here. Details of these techniques are described in such books as Monoclonal Antibodies-Hybridomas: A New Dimension in Biological Analysis, edited by Roger H. Kennett et al., published by Plenum Press (1980).

Hybridomas producing monoclonal antibodies specific for epitopes which are found on the A4 28-mer peptide are most effectively produced by first immunizing an animal from which hybridomas can be produced such as, for example, a Balb/c mouse, with initial subcutaneous injections of the 28-mer peptide in Freund's adjuvant, followed by booster injections within a few days. The fusion can be carried out using any of the techniques commonly known to those of ordinary skill in the art. The screening of the hybridomas to determine which ones are producing monoclonal antibodies specific for the 28-mer peptide is straightforward and can be done either in a standard ELISA or RIA format. For example, in an RIA screening format the culture supernatant, or ascites fluid from a hybridoma producing monoclonal antibody is reacted with ¹²⁵I-28-mer peptide.

The antibody of the present invention can be utilized in immunoassays for the detection of A4-amyloid polypeptide wherever it may occur, including fluid or semi-fluid human samples. The immunoassays can be competitive or sandwich, as is otherwise well known they all depend on the formation of antibody-antigen immune complex. These assays are not described herein in any further detail, as they are well known to those of skill in the art.

For purposes of the assays, the antibody can be immobilized or labeled.

There are many carriers to which the antibody can be bound for immobilization and which can be used in the present invention. Well-known carriers include glass,

polystyrene, polypropylene, polyethylene, dextran, nylon, amylases, natural and modified celluloses, polyacrylamides, agaroses, and magnetite. The nature of the carrier can be either soluble to some extent or insoluble for purposes of the invention. Those skilled in the art will know many other suitable carriers for binding the antibody, or will be able to ascertain such, using routine experimentation.

Depending on the particular embodiment of the invention, the antibody will be coupled with a detectable label such as an enzyme, radioactive isotope, fluorescent compound, chemiluminescent compound, or bioluminescent compound.

Those of ordinary skill in the art will know of other suitable labels for binding to the antibody or will be able to ascertain such using routine experimentation. Furthermore, the binding of these labels to the antibody can be done using standard techniques commonly known to those of ordinary skill in the art.

The antibody can be bound to an enzyme. This enzyme, in turn, when later exposed to its substrate will react to the substrate in such a manner as to produce a chemical moiety which can be detected, as, for example, spectrophotometric or fluorometric means. Examples of enzymes that can be used to detectably label are malate dehydrogenase, staphylococcal nuclease, delta-5-steroid isomerase, yeast alcohol dehydrogenase, alpha-glycerophosphate dehydrogenase, triose phosphate isomerase, alkaline phosphatase, asparaginase, glucose oxidase, beta-galactosidase, ribonuclease, urease, catalase, glucose-6-phosphate dehydrogenase, glucoamylase, and acetylcholine esterase.

The presence of an antibody can also be detected by labeling it with a radioactive isotope. The presence of the radioactive isotope could then be determined by such means as the use of a gamma counter or a scintillation counter. Isotopes which are particularly useful are ³H, ¹²⁵I, ³²P, ³⁵S, ¹⁴C, ⁵¹Cr, ³⁶Cl, ⁵⁷Co, ⁵⁸Co, ⁵⁹Fe, ⁷⁵Se, and ¹⁵²Eu.

It is also possible to detect the presence of the antibody by labeling it with a fluorescent compound. When the fluorescently labeled antibody is exposed to light of the proper wavelength, its presence can then be detected due to fluorescence of the dye. Among the most important fluorescent labeling compounds are fluorescein isothiocyanate, rhodamine, phycoerythrin, phycocyanin, allophycocyanin, o-phthaldehyde, and fluorescamine.

Another way in which the antibody can be detectably labeled is by coupling it to a chemiluminescent compound. The presence of the chemiluminescent-tagged antibody is then determined by detecting the presence of luminescence that arises during the course of a chemical reaction. Examples of particularly useful chemiluminescent labeling compounds are luminol, isoluminol, aromatic-acridinium ester, imidazol, acridinium salt, and oxalate ester.

Likewise, a bioluminescent compound may also be used to label the antibody. Bioluminescence is a special

type of chemiluminescence which is found in biological systems and in which a catalytic protein increases the efficiency of the chemiluminescent reaction. The presence of a bioluminescent binding partner would be determined by detecting the presence of luminescence. Important bioluminescent compounds for purposes of labeling are luciferin, luciferase, and aequorin.

The antibody for use in the assay of the invention are ideally suited for the preparation of a kit. Such a kit may comprise a carrier means being compartmentalized to receive in close confinement one or more container means such as vials, tubes, and the like, each of said container means comprising one of the separate elements to be used in the method.

For example, one of the container means may comprise a first antibody bound to an insoluble or partly soluble carrier. A second container may comprise soluble, detectably-labeled second antibody, in lyophilized form or in solution. The carrier means may also contain a third container means comprising a detectably-labeled third antibody in lyophilized form or in solution. Such a kit can be used for sandwich assays. See, e.g., David *et al.* USP 4, 376, 110 herein incorporated by reference.

In addition, the carrier means may also contain a plurality of containers each of which comprises different, predetermined amounts of known A4-amyloid antigen. These latter containers can then be used to prepare a standard curve into which can be interpolated the results obtained from the sample containing the unknown amount of A4-amyloid antigen.

Imaging can be carried out *in vitro* or *in vivo*. *In vitro* imaging can be done with the labels mentioned previously. *In vivo* imaging is done with diagnostically effective labeled antibodies. The term "diagnostically effective" means that the amount of detectably labeled antibody administered is sufficient to enable detection of the site of amyloid presence when compared to a background signal.

diagnostic imaging can be utilized in accordance with this invention.

Another important factor in selecting a radionuclide for *in vivo* diagnosis is that the half-life of a radionuclide be long enough so that it is still detectable at the time of maximum uptake by the target, but short enough so that deleterious radiation upon the host is minimized. Ideally, a radionuclide used for *in vivo* imaging will lack a particulate emission, but produce a large number of photons in a 140-200 keV range, which may be readily detected by conventional gamma cameras.

For *in vivo* diagnosis, radionuclides may be bound to antibody either directly or indirectly by using an intermediary functional group. Intermediary functional groups which are often used to bind radioisotopes which exist as metallic ions to antibody are diethylenetriaminepentaacetic acid (DTPA) and ethylenediaminetetraacetic acid (EDTA). Typical examples of metallic ions which can be bound to immunoglobulins are ^{99m}Tc , ^{123}I , ^{111}In , ^{131}I , ^{97}Ru , ^{67}Cu , ^{67}Ga , ^{125}I , ^{68}Ga , ^{72}As , ^{89}Zr , and ^{201}Tl .

The antibody used in the method of the invention can also be labeled with paramagnetic isotopes for purposes of *in vivo* diagnosis. Elements which are particularly useful (as in Magnetic Resonance Imaging (MRI) techniques) in this manner include ^{157}Gd , ^{55}Mn , ^{162}Dy , ^{52}Cr , and ^{56}Fe .

Preparations of the imaging antibody for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, and emulsions. Examples of non-aqueous solvents are propyleneglycol, polyethyleneglycol, vegetable oil such as olive oil, and injectable organic esters such as ethyloleate. Aqueous carriers include water, alcoholic/aqueous solutions, emulsions or suspensions, including saline and buffered media, parenteral vehicles including sodium chloride solution, Ringer's dextros, dextrose and sodium chloride, lactated Ringer's, or fixed oils. Intravenous vehicles include fluid and nutrient electrolyte replacement,

Preparation and Characterization of the Synthetic Amyloid Polypeptide

The A4 amyloid polypeptide of 28 residues (Fig. 1), corresponding to the previously reported sequence of Masters *et al.* (1985a), was synthesized on a Bioscience SAM2 synthesizer using the general procedure of Merrifield (1963). Purification was achieved with a 3 X 65 cm column of Sephadex G50 (10-40 μ). Aliquots were removed, spotted onto TLC plates and sprayed with fluorescamine to locate protein. Material was pooled and loaded onto an analytical HPLC column (Vydac C₁₈, catalog No. 218TP54). Elution was carried out at a flow rate of 1.7 ml/min. with 0.05% trifluoroacetic acid/H₂O for 5 min followed by a 5-100% linear gradient of 0.05% TFA/CH₃CH for 17 min. The optical density profile at 230 nm revealed a single major peak that was further analyzed. Amino acid analysis was carried out in 6 M HCl containing 1% phenol for 18 hours at 110°C. The sample was dried under N₂, dissolved in citrate buffer and analyzed with an LKB 4151 Alpha Plus amino acid analyzer. Amino acid analyses were consistent with the published sequence and indicated an approximate yield of 95-98%. Peptide samples were stored as a dry powder at -20°C until used. For some experiments, the peptide was linked to Kehole Limpet Hemocyanin (BABCO, Berkeley, CA) prior to use.

Preparation of Polyclonal Antibodies (Pabs)

New Zealand white female rabbits were used for the production of polyclonal antibodies to the synthetic amyloid peptide. In some cases AP linked to KLH was used as the antigen. Subdermal injection was carried out using 1 mg of AP that had been emulsified in Freund's complete adjuvant. After 3 weeks the animals were bled and tested for reactivity. Animals were injected again after 3 weeks using 1 mg of AP in Freund's incomplete adjuvant. Two weeks later the serum was tested and was observed to give a positive reaction at 1/1,000 dilution. Rabbits were then injected with 1 mg AP for 2 monthly intervals after which the serum was positive at a dilution of greater than 1/10,000 when assayed by immunoblotting.

Preparation of Monoclonal Antibodies (Mabs)

Balb/c mice were injected subdermally with 1 mg each of AP in Freund's complete adjuvant. After 3 weeks sera were positive at a dilution of 1/1,000 using the

been deposited before the filing date of the present application at the American Type Culture Collection, Rockville, Maryland, under the terms of the Budapest Treaty and given accession number HB9542.

Dot Blot Assay

Antibodies were tested for reactivity to AP using the BioRad dot blot apparatus according to the manufacturer's directions. For initial screening, 1 mg of AP was sonicated in 0.5 ml of 1% sodium dodecylsulfate in H₂O and added to an equal volume of 2.5% Triton X-100, 0.3 M NaCl, 40 mM Tris HCl, pH 7.4. One μ g AP was added to each well followed by 50 μ l of 10% BSA. For Mab assays, 150 μ l of culture supernatant was added per well. Pab assays used serum diluted 1/500, 1/1,000 and 1/10,000 in 150 μ l Tris buffered saline (TBS) containing 0.15 M sodium chloride, 20 mM Tris, pH 7.4. After filtration TBS was used for intermediate washes between antibody additions and prior to adding substrate. 50 μ l of horseradish peroxidase-conjugated affinity purified, goat anti-mouse or anti-rabbit IgG (Cappel), diluted 1/2,000 in 5% BSA, 0.5% Triton X-100, 0.15 M NaCl, 20 mM Tris hydrochloride, pH 7.4 was added to each well. The reaction product was visualized using diaminobenzidine, 0.5 mg/ml, imidazole, 1 mg/ml, and H₂O₂, 0.015%. Negative controls consisted of tissue culture media or pre-immune sera, omission of AP, and addition of a monoclonal antibody supernatant specific for a protein other than AP. For subsequent assays, the amount of antigen per well was varied over the range 0.001-1 μ g and antibodies were tested at different dilutions. For these assays, individual strips of nitrocellulose containing various amounts of antigen were immunostained as described by Brown *et al.* (1983).

PAGE Procedures

PAGE procedures were carried out as described previously (Brown *et al.*, 1981, 1982). Variations from the published procedures are described in the text.

Immunoblot Assay

Electrophoretic transfer of proteins to nitrocellulose membranes and immunostaining procedures were carried out as previously described (Brown *et al.*, 1983).

Immunohistology

Thioflavin S Staining

Formalin-fixed sections were placed in a 0.1% solution of Thioflavin S (Sigma) in TBS for 10 min. Excess stain was removed by placing the tissue sections in 70% ethanol for 1 to 2 min and then in water. Sections were coverslipped using a solution containing 20% polyvinyl alcohol (Sigma), 10% glycerol and 50 mM Tris HCl, pH 8.5. Where indicated in the text, double staining was carried out on the same tissue: a section stained with Thioflavin S was subsequently stained by means of the immunocytochemical procedure described above.

Computer-Assisted Image Enhancement

Specimens of prefrontal cortex immunostained for amyloid protein were visualized through a Leitz Laborlux 12 light microscope equipped with a MCI 65 televideo camera that interfaced with a computer. The transmitted images of immunostained amyloid deposits were processed by the computer using software equipped with pseudocolor optical density coding. The transmitted image was reconstructed on a separate monitor according to the individual colors assigned to each level of gray in the image. Thus, internal density variations within amyloid deposits could be assessed.

RESULTS

Peptide Preparation and Characterization

A synthetic amyloid peptide (AP) of 28 amino acids (Fig. 1), with a calculated molecular weight of 3.2 kD, was synthesized and then analyzed by PAGE procedures prior to immunological studies. The AP was dissolved in PAGE sample buffer containing 2% SDS, 5% mercaptoethanol, and 9.5 M urea and was electrophoresed on a 10% gel containing 0.1% SDS. After staining with Coomassie blue the peptide appeared as a broad band at approximately 23-25 kD and a narrow band that migrated at the gel front during electrophoresis. The higher molecular weight species appeared to be an aggregate since it was eliminated by adding urea to the separating gel: the AP was dissolved in sample buffer containing 9.5 M urea and electrophoresed on a 10% or 15% gel that contained 6 M urea and 2% SDS. After staining with Coomassie blue, the predominant species appeared as a 3-4 kD band which is consistent with the

Using a rapid dot blot screening procedure, it was observed that antiserum Scarlet-1 prepared to the unconjugated polypeptide was reactive at the 0.01 µg level and produced staining above background with 0.001 µg of antigen. At this antigen level, the reaction product from pre-immune serum was barely observable. Scarlet-1 was more potent than serum from a rabbit immunized with the KLH-linked derivative of AP.

Scarlet-1 was characterized with respect to the state of aggregation of AP. Although the dot blot assay may have contained primarily aggregated complexes of the polypeptide, this possibility was examined further by PAGE. AP was electrophoresed in a gel containing 2% SDS under conditions that allowed aggregates to form. An electroblot was prepared from the same gel and immunostained with Scarlet-1. The antiserum detected the lower molecular weight form as well as aggregated species. Thus, antiserum produced in response to the amyloid polypeptide was extremely potent irrespective of the state of aggregation of AP.

Characterization of Monoclonal Antibodies (Mabs)

The supernatants of 24 hybridomas were positive for AP as indicated by the dot blot assay. Of these, three Mabs with particularly strong binding properties were further characterized. Dot blots demonstrated that the three Mabs, designated 4E12, 5E2 and 10H3, were at least as reactive as the polyclonal antisera. In some immunocytochemical experiments (see below), a mixture of the three Mabs in equal parts was used since the mixed preparation was also intensively reactive towards the synthetic peptide. As with polyclonal sera, each of the Mabs reacted strongly with the AP that had been previously treated with SDS prior to electrophoresis.

Immunodetection of Amyloid with Polyclonal Antisera

Initial immunostaining studies employed polyclonal antisera to AP; the data obtained served as a basis for comparison with later experiments using Mabs. Prefrontal cortex (PC) of an AD case contained numerous plaques and intraneuronal NFTs that were readily visualized by staining with Thioflavin S (Kelenyi, 1967). Immunostaining the same tissues with Scarlet-1 clearly revealed amyloid plaques that were typical of those seen in the PC and hippocampus of four AD cases.

The same results were obtained with antisera prepared

material in the parenchyma with the vascular amyloid. The double label technique applied to blood vessels cut in cross-section confirmed that all vascular layers that bound Thioflavin S were detectable with Scarlet-1 immune serum.

Monoclonal antibodies were prepared to the A4 28 amino acid polypeptide derived from AD brain amyloid (Masters *et al.*, 1985a, Figure 1). The supernatants of 24 hybridomas were positive; of these, three Mabs with particularly strong binding properties were further characterized. Dot blots demonstrated that the three Mabs, designated 4E12, 5E2 and 10H3, were strongly reactive towards the polypeptide even at extremely high dilutions. In some immunocytochemical experiments (see below) a mixture of the three Mabs in equal parts was used.

Initial studies were aimed at establishing the specificity of the Mabs by immunostaining AD brain sections using an avidin-biotin horseradish peroxidase procedure and analyzing the epitopic distribution by conventional imaging methods. Sections of prefrontal cortex were used. A section stained with thioflavin S, which is known to react with brain amyloid, was counter-stained with Mab 5E2. The antibody bound to the amyloid deposit with a distribution that overlapped the fluorescent dye. In other studies, observations on numerous plaques stained with the three Mabs indicated that these antibodies provided more detailed architectonic information than previously reported for AD brain amyloid deposits. The Mabs demonstrated that target epitopes occurred in deposits of different sizes and different morphologies. For example, it was observed that the 10H3 epitope was localized both within a core and in a peripheral ring whereas in other instances the same epitope was more randomly distributed. Immunodetection of an amyloid core surrounded by a ring during the course of these studies was an original finding as were other morphologies described below.

Although it is not known whether or not the three Mabs under consideration are specific for the same or different epitopic sites of the A4 28-mer amyloid polypeptide sequence, when mixed together they provided particularly intense staining of tissues; however, the dark reaction product remained in sharp contrast to low background staining. In control studies, there was no staining with the Mab mixture applied to neurologically normal controls beyond light background staining. The specific and intense staining of amyloid with the mixed Mabs appeared to be the method of choice for more detailed

dures that provided increased resolution of structural features. Sections of prefrontal cortex immunostained with the Mab mixture were viewed by means of a light microscope equipped with a televideo camera that interfaced with a computer for the production of processed images. The images were digitized, size estimates were made, and pseudocolor gray scaling was used to display different levels of density. The major types of amyloid deposits were distinguished in terms of size, internal organization, and internal density.

Four classes of amyloid deposits of different sizes were identified. Small punctate amyloid deposits ($9.06 \pm 0.24 \mu\text{m}$ diameter) were the most commonly observed immunostained configuration. Often seen adjacent to punctate deposits were minute amounts of material that may represent precursor forms. Amyloid accumulations referred to as macular amyloid deposits ($30.87 \pm 1.28 \mu\text{m}$ diameter) were considered together as a class since they are distinguished by a larger diameter. In one example, multiple foci of dense deposits were present throughout the field; a second macular structure of similar size contained only diffuse reaction product. Another example appeared as a darker staining accumulation of amyloid.

Also observed with ring-like amyloid deposits ($40.51 \pm 4.65 \mu\text{m}$ diameter) in which the central region contained little or no immunodetectable amyloid. The rarest configuration was a ring + core amyloid deposit (a ring of amyloid that contained a distinct and separate amyloid core, the ring measured $48.73 \pm 7.36 \mu\text{m}$ diameter and the core measured $12.85 \pm 2.20 \mu\text{m}$ diameter). Previous immunostaining of amyloid deposits with polyclonal sera to a synthetic polypeptide did not detect the ring or ring with core patterns (Master *et al.*, 1985b; Wong *et al.*, 1985).

With respect to neocortex, punctate deposits were predominantly located in layer I and their frequency of occurrence was inversely related to their depth within the cortical mantle. By contrast, macular deposits had a different distribution. When compared with punctate deposits, the macular types were represented to a lesser extent in layer I, but to a greater degree in layers II through VI. Ring shapes occurred in all layers. Ring with core structures were less frequently encountered than the other morphologies. This laminar distribution of the different amyloid classes has not been previously described.

The immunoperoxidase-stained amyloid was subjected to pattern analysis by computer-enhanced imag-

deposits of similar size and internal heterogeneity. Irrespective of the overall morphologic variations among the four groups of amyloid deposits, a common feature was the presence of multiple foci of high density.

It is to be emphasized that the Mabs used in the present study exhibited high specificity for amyloid and not necessarily for the senile plaque detectable by silver staining methods. Due to this immunologic specificity, observations were not made with regard to senile plaques per se, which include a variety of cellular and subcellular elements in addition to amyloid (Wisniewski and Terry, 1973); instead, our attention was focused upon morphologic entities identifiable by Mabs with high specificity towards amyloid. In other studies, we have observed that Bielchowsky staining demonstrates that layers II and III contain the greatest number of senile plaques while not revealing the punctate lesions of layer I or other layers in similar tissue sections. For this reason, it is important to emphasize that there is only partial overlap between the immunodetectable amyloid deposit and the classically defined senile plaque.

While the macular amyloid deposits may correspond to the classical descriptions of amyloid within senile plaques (Wisniewski and Terry, 1973), the remaining forms we described appear to be unique. Specifically, Mabs to the A4 28-mer polypeptide visualized a series of amyloid deposits in the AD brain that do not appear to have been previously described: punctate, ring and ring+core amyloid deposits that each have internal density gradients are original findings. The unique aspect of these Mabs suggests their use as powerful reagents for the detailed investigation of subtypes of AD; such reagents have not been previously available.

The techniques discussed above appear well-suited to the production of the Mab of the invention which is unique and which provides a new tool to examine the molecular pathogenesis of AD.

REFERENCES

- Allsop, D., et al., Brain Res. 259:348-352 (1983).
 Brown, B.A., et al., J. Neurochem. 40:299-308 (1983).
 Brown, B.A., et al., J. Neurochem. 36:143-153 (1981).
 Brown, B.A., J. Cell Biol. 94:159-164 (1982).
 Divry, P., J. Neurol. Psychiat. 27:643-657 (1927).
 Merz, P.A., et al., Acta Neuropathol. (Berl.) 60:113-124 (1983).
 Wisniewski, H.M., et al., Ann. N.Y. Acad. Sci. 396:119-129 (1982).
 Wisniewski, H.M., et al., "Reexamination of the pathogenesis of the senile plaque," in H.M. Zimmerman (ed.), Progress in Neuropathology, pp. 1-26, Grune and Stratton, New York (1973).
 Wong, W.C., et al., Proc. Natl. Acad. Sci. U.S.A. 82:8729-8732 (1985).
- ### Claims
1. A hybridoma deposited under the accession number ATCC HB9542.
 2. An antibody expressed by the hybridoma designated 10H3, deposited under the accession number ATCC HB9542.
 3. An antibody as claimed in claim 2 which is detectably labelled.
 4. An antibody as claimed in claim 3 wherein the detectable label is a radiolabel, an enzyme label, a cofactor label, a chemiluminescent label, a bioluminescent label, a fluorescent label, a paramagnetic label, or a metal label.
 5. An antibody as claimed in claim 2 which is bound to a polymeric carrier.
 6. An immunoassay method for the determination or detection of A4-amyloid in a sample, the method comprising contacting a sample with an antibody to A4-amyloid as claimed in claim 2 and determining whether an immune complex is formed between the antibody and the A4-amyloid.
 7. A method for the detection of the presence of A4-amyloid in a tissue sample, the method comprising:
 - obtaining a tissue sample from the body of a subject;
 - contacting the tissue sample with an imaging-effective amount of a detectably labelled antibody as

detecting the label to determine the in vivo distribution of A4-amyloid.

10. The use of an antibody as claimed in any of claims 2 to 5 in the preparation of an agent for the diagnosis of Alzheimer's disease. 5
11. A diagnostic composition comprising an antibody as claimed in any of claims 2 to 5 and a pharmaceutically acceptable carrier. 10
12. An immunoassay kit for the diagnosis of Alzheimer's disease comprising an antibody as claimed in any of claims 2 to 5 and carrier means for the antibody. 15
13. An antibody as claimed in claim 3 or claim 4 for use in a method for the detection of the presence and/or distribution of A4-amyloid in one or more tissues, wherein the detection is carried out by imaging in vivo, the method comprising: 20
 - administering an imaging-effective amount of the antibody, which is detectably labelled, and detecting the label to determine the in vivo presence and/or distribution of A4-amyloid.

Patentansprüche

1. Hybridom hinterlegt unter der Zugriffsnummer ATCC HB90542. 25
2. Antikörper, der durch das als 10H3 bezeichnete Hybridom exprimiert wird, das unter der Zugriffsnummer ATCC HB9542 hinterlegt ist. 30
3. Antikörper nach Anspruch 2, welcher auffindbar markiert ist. 35
4. Antikörper nach Anspruch 3, bei welchem die auffindbare Markierung ein Radiomarker, ein Enzymmarker, ein Cofaktormarker, ein Chemilumineszenzmarker, ein Biolumineszenzmarker, ein Fluoreszenzmarker, ein paramagnetischer Marker oder ein Metallmarker ist. 40
5. Antikörper nach Anspruch 2, welcher an einen polymeren Träger gebunden ist. 45
6. Immunoassayverfahren zur Bestimmung oder Auffindung von A4-Amyloid in einer Probe, welche Methode das Kontaktieren der Probe mit einem Antikörper gegen A4-Amyloid gemäß Anspruch 2 und das Bestimmen, ob ein Immunkomplex zwischen dem Antikörper und dem A4-Amyloid gebildet ist, umfaßt. 50
7. Verfahren zur Bestimmung der Präsenz von A4-Amyloid in Gewebeproben, wobei das Verfahren

Körper einer Person;

Kontaktieren der Gewebeprobe mit einer anzeigefähigen Menge eines auffindbar markierten Antikörpers gemäß Anspruch 3 oder 4; und

Ermittlung der Markierung zur Feststellung der Präsenz von A4-Amyloid in der Gewebeprobe.

8. Verfahren nach Anspruch 7 wobei die Gewebeprobe eine Gewebesektion ist.
9. Verfahren zur nichtdiagnostischen Ermittlung der Präsenz oder Verteilung von A4-Amyloid in einem oder mehreren Geweben, wobei die Ermittlung durch in vivo Anzeige durchgeführt wird, welches Verfahren umfaßt:
 - Verabreichung einer anzeigefähigen Menge eines auffindbar markierten Antikörpers gemäß Anspruch 3 oder 4; und
 - Ermittlung der Markierung zur Bestimmung der in vivo Verteilung von A4-Amyloid.

10. Verwendung eines Antikörpers nach einem der Ansprüche 2 bis 5, bei der Herstellung eines Mittels gegen die Alzheimer'sche Krankheit.

11. Diagnosezusammensetzung, welche einen Antikörper nach einem der Ansprüche 2 bis 5 und einen pharmazeutisch verträglichen Träger aufweist. 25

12. Immunoassay-Ausrüstung zur Diagnose der Alzheimer'schen Krankheit, welche Ausrüstung einen Antikörper nach einem der Ansprüche 2 bis 5 und einen Träger für den Antikörper aufweist. 30

13. Antikörper nach Anspruch 3 oder 4 zur Verwendung in einem Verfahren zur Ermittlung der Präsenz und/oder Verteilung von A4-Amyloid in einem oder mehreren Geweben, wobei die Ermittlung durch in vivo Anzeige durchgeführt wird, wobei das Verfahren umfaßt: 35

Verabreichung einer anzeigefähigen Menge des Antikörpers, der auffindbar markiert ist, und Ermittlung der Markierung, um die in vivo Präsenz und/oder Verteilung von A4-Amyloid zu bestimmen. 40

Revendications

1. Hybridome déposé sous le numéro d'accès ATCC HB9542. 45
2. Anticorps exprimé par l'hybridome appelé 10H3, déposé sous le numéro d'accès ATCC HB9542.
3. Anticorps suivant la revendication 2, qui est marqué de manière détectable. 50

- marqueur enzymatique, un marqueur de type cofacteur, un marqueur chimioluminescent, un marqueur bioluminescent, un marqueur fluorescent, un marqueur paramagnétique ou un marqueur métallique.
- 5
5. Anticorps suivant la revendication 2, qui est fixé à un support polymère.
6. Procédé d'immunodosage pour la détermination ou la détection de l'amyloïde de séquence A4 dans un échantillon, le procédé comprenant la mise en contact d'un échantillon avec un anticorps de l'amyloïde de séquence A4 suivant la revendication 2 et la détermination de la formation ou non d'un complexe immun entre l'anticorps et l'amyloïde de séquence A4.
- 10
- 15
7. Procédé pour la détection de la présence de l'amyloïde de séquence A4 dans un échantillon tissulaire, le procédé comprenant :
- 20
- le prélèvement d'un échantillon tissulaire du corps d'un sujet;
- la mise en contact de l'échantillon tissulaire avec une quantité efficace pour l'imagerie d'un anticorps marqué de manière détectable suivant la revendication 3 ou 4, et
- 25
- la détection du marqueur pour établir la présence de l'amyloïde de séquence A4 dans l'échantillon tissulaire.
- 30
8. Procédé suivant la revendication 7, dans lequel l'échantillon tissulaire est une coupe de tissu.
9. Procédé pour la détection non diagnostique de la présence ou de la distribution de l'amyloïde de séquence A4 dans un ou plusieurs tissus, dans lequel la détection est réalisée par imagerie in vivo, le procédé comprenant :
- 35
- l'administration d'une quantité efficace pour l'imagerie d'un anticorps marqué de manière détectable suivant la revendication 3 ou 4, et
- 40
- la détection du marqueur pour établir la distribution in vivo de l'amyloïde de séquence A4.
10. Utilisation d'un anticorps suivant l'une quelconque des revendications 2 à 5, dans la préparation d'un agent pour le diagnostic de la maladie d'Alzheimer.
- 45
11. Composition diagnostique comprenant un anticorps suivant l'une quelconque des revendications 2 à 5 et un support pharmaceutiquement acceptable.
- 50
12. Kit d'immunodosage pour le diagnostic de la maladie d'Alzheimer, comprenant un anticorps suivant l'une quelconque des revendications 2 à 5 et un dispositif support pour l'anticorps.
- 55
13. Anticorps suivant la revendication 3 ou 4, à utiliser dans un procédé pour la détection de la présence

et/ou de la distribution de l'amyloïde de séquence A4 dans un ou plusieurs tissus, dans lequel la détection est réalisée par imagerie in vivo, le procédé comprenant :

l'administration d'une quantité efficace pour l'imagerie d'un anticorps marqué de manière détectable, et

la détection du marqueur pour déterminer la présence et/ou la distribution in vivo de l'amyloïde de séquence A4.

FIG.1

β -polypeptide: N-asp-ala-glu-phe-arg-his-asp-ser-gly-tyr-
 11 24
 gln-val-his-his-gln-lys-leu-val-phe-phe-ala-glu-asp-val-C

A4: N-asp-ala-glu-phe-arg-his-asp-ser-gly-tyr-glu-val-his-
 11 27 28
 his-gln-lys-leu-val-phe-phe-ala-glu-asp-val-gly-ser-ser-ala-C

USP: N-asp-ala-glu-phe-arg-his-asp-ser-gly-tyr-gln-val-his-
 11 27 28
 his-gln-lys-leu-val-phe-phe-ala-glu-asp-val-gly-ser-asn-lys-C